

strong evidence that non-competitive binding sites exist at non- α interfaces of heteromeric nAChRs. Furthermore, non-competitive antagonist binding of natural alkaloids has been shown to occur at the channel pore. In previous studies using Analogue 1, the ion channel pore appears to be the main binding site for this compound. In contrast Analogue 2 does not bind to this site. Here, we report studies of the non-competitive binding site within the N-terminal domain on the $\alpha 4\beta 2$ nAChR. Since this $\beta(-)\alpha(+)$ subunit interface of the N-terminal domain is not well studied, water accessibility of the residues was first examined using the Substituted Cysteine Accessibility Method (SCAM). The residues on Loop D (N88, V89, W90, V91, K92, Q93 and E94) of the $\alpha 4$ subunit and Loop A (V116, V117, L118, Y119, N120, N121, A122, D123 and G124) of the $\beta 2$ subunit were individually mutated to cysteine, expressed in *Xenopus* oocytes and analysed using two-electrode voltage clamp recordings. Surface accessibility was tested by evaluating the reaction of sulphhydryl reagent ethylammonium-methanethiosulfonate (MTSEA) in the opened (in the presence of ACh) and closed channel states (in the absence of ACh). The site was then evaluated using two methods: (1) The antagonists were competed with the sulphhydryl reagents where protection from irreversible inhibition infers the binding site. (2) Analogue 1 and 2 were synthesized into a thiol reactive probe capable of reacting with cysteine directly. Irreversible inhibition infers the binding site. All mutants generated functional receptors and most were accessible to MTSEA. Both competition and reactive probe experiments showed that neither of these analogues bind within the N-terminal domain. Other loops within the non-competitive $\beta(-)\alpha(+)$ interface and the competitive $\alpha(+)\beta(-)$ interface will be studied in the future.

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Mutation of proline enables subtype selectivity of α -conotoxin BuIA

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α -Conotoxins are neuroactive peptides, isolated from the venom of carnivorous snails that act as competitive antagonists of nicotinic acetylcholine receptors (nAChRs). α -Conotoxins are small peptides that have two cysteine loops and a highly conserved proline (Pro) in the first loop. Crystal structures of α -conotoxins in complex with the acetylcholine binding protein (AChBP) show that the α -conotoxin Pro side chain is positioned to potentially interact with the ACh binding pocket. BuIA is a 13 amino acid α -conotoxin that kinetically discriminates between $\beta 2$ - and $\beta 4$ -containing nAChRs; the off-rate of BuIA is slow for all $\beta 4$ - vs. $\beta 2$ -containing nAChRs. Three residues on the β subunit at positions 59, 111 and 119 are critical for binding of some α -conotoxins. These residues line the putative acetylcholine-binding pocket and differ between $\beta 2$ and $\beta 4$ nAChR subunits. Site-directed mutagenesis has demonstrated that Thr59 is an important determinant of sensitivity for α -conotoxins as well as other competitive antagonists. Homology modeling studies with the AChBP identified Val111 and Phe119 as likely residues interacting with α -conotoxins MII and PnIA. BuIA contains two Pro residues. In the present study we explored the role of the BuIA Pro residues in the ability of BuIA to discriminate between $\beta 2$ - and $\beta 4$ -containing nAChRs. We hypothesized that Pro6 and/or Pro7 interacts with non-conserved

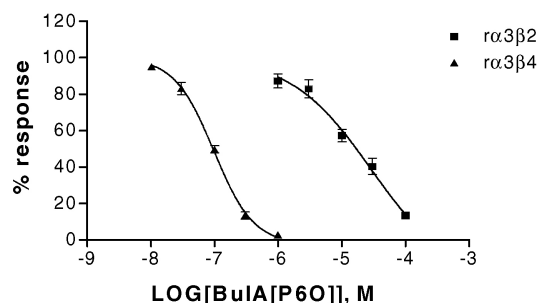


Fig. 1. Presence of hydroxyproline enables α -conotoxin BuIA to discriminate between $\alpha 3\beta 2$ and $\alpha 3\beta 4$ nAChRs.

residues on the nAChR β subunit and through this interaction influences the subtype selectivity of BuIA. BuIA as well as BuIA analogs were synthesized using Fmoc chemistry. Pro6 or Pro7 was substituted with 4-*trans* hydroxyproline or 3-*trans* hydroxyproline. nAChR residues present in the $\beta 4$ subunit were introduced as point mutations in the homologous positions in the $\beta 2$ subunit. These mutations included $\beta 2T59K$, $\beta 2V111I$ and $\beta 2F119Q$. In addition, one mutant $\beta 4$ subunit was made, $\beta 4K59T$. Two-electrode voltage clamp of oocytes injected with cRNA encoding wild type and mutant nAChRs was used to assess the activity of the conotoxin analogs. The interaction between the α -conotoxin BuIA analogs and the β subunit of the nAChR was assessed by double-mutant cycle analysis; pair-wise interaction energies of Pro6 and Pro7 with nAChR residues (at positions 59, 111 and 119) were determined. Pro6 interacts with Thr59 (on the $\beta 2$ subunit) with a coupling energy of 2.4 kcal/mol and Pro6 interacts with Lys59 (on the $\beta 4$ subunit) with a coupling energy of 2.6 kcal/mol (energies are absolute values). The introduction of 4-*trans* hydroxyproline in the 6th position selectively decreased binding of BuIA to $\alpha 3\beta 2$ nAChRs thus enabling BuIA to selectively block $\alpha 3\beta 4$ vs. $\alpha 3\beta 2$ nAChRs (Fig. 1). Pro6 thus represents an amino acid that may be mutated to create α -conotoxins with improved subtype selectivity.

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Isoanatabine, a naturally occurring $\alpha 4\beta 2$ nicotinic receptor agonist

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Anabaseine was the first nemertine alkaloid to be isolated and pharmacologically characterized; benzylidene-substituted anabaseines including DMXBA (GTS-21) are being investigated as potential therapeutic agents for treating cognitive dysfunction (Kem, 2000; Freedman et al., 2008). Here we examine the pharmacological properties of isoanatabine [2-(3-pyridyl)-1,2,5,6-tetrahydropyridine], an anabaseine isomer, that was isolated from a different species of nemertine. Enantiomers of synthetic isoanatabine and anatabine were obtained by chiral HPLC. Functional properties (EC_{50} and I_{max}) were assessed on *Xenopus* oocytes ($n \geq 4$) using 100 μM ($\alpha 4\beta 2$) or 1000 μM ($\alpha 7$) ACh as standards; $\alpha 4\beta 2$ nAChR binding was measured by displacement of [3H]-cytisine using rat brain membranes. Data were fitted with Prism software, to yield calculated properties shown below:

Rat $\alpha 4\beta 2$	Human $\alpha 4\beta 2$		Human $\alpha 7$ (Enant)-compound		
Ki (nM)	EC ₅₀ (μ M)	I _{max} (%ACh)	EC ₅₀ (μ M)	I _{max} (%ACh)	
(S)-Isoanatabine	650	0.91	78.8	45.1	76.1
(R)-Isoanatabine	198	0.32	103	51.9	31.2
(S)-Anatabine	282	0.79	39.8	33.0	103
(R)-Anatabine	114	0.48	17.2	41.1	90.3
(S)-Nicotine	5.6	0.21	84.3	56.5	91.7
(S)-Anabasine	1,100	7.91	76.0	18.4	100
Anabaseine	94	~12	~8	18	100

(R)-Isoanatabine (1) is 3-fold more potent than (S)-isoanatabine at $\alpha 4\beta 2$ receptor, but its $\alpha 7$ I_{max} is only about half as great; (2) is a potent $\alpha 4\beta 2$ agonist comparable to (S)-nicotine; (3) displays a much higher efficacy at the $\alpha 4\beta 2$ receptor and a much lower I_{max} at $\alpha 7$ relative to anatabine; (4) relative to anabasine, is a 9-fold more potent $\alpha 4\beta 2$ partial agonist but a 3-fold less efficacious $\alpha 7$ partial agonist; (5) Isoanatabine, relative to anabaseine, is a much more potent and efficacious $\alpha 4\beta 2$ agonist. The 3,4 position double bond in isoanatabine and, to a lesser extent, the 4,5-position double bond in anatabine contribute greatly to their $\alpha 4\beta 2$ agonist activities.

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Automated two-electrode voltage clamp for medium-throughput studies of ion channels with non-destructive sample analysis

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Electrophysiological recordings using patch clamp systems are important tools for the study of ion channels and receptors. Although conventional patch clamp delivers high-quality data, it does not allow a fast high-throughput screening of drugs on ion channels or receptors, and is mostly limited to record data of maximum 40 cells daily by a skilled scientist. Here, we present a new automated two-electrode voltage clamp, which allows a fast, reliable, and high-quality screening of up to 96 drugs in a single experiment. Our system is equipped with one 96-well plate, which contains the samples (or toxins) using minimal volumes (220 μ l). A fact that is especially important when probing effects of samples that are only available in minute amounts such as toxins. Since the automate uses a non-destructive measurement with drugs stored in the 96-well plate, samples can be reused several times to evaluate their effects either at the same channel subtype or to another membrane protein. A second 96-well plate contains the injected oocytes from *Xenopus*. Oocytes are automatically loaded and poked and their properties assessed to determine the membrane quality. As the automate works fully unattended, measurements can be carried continuously over 24 h. Depending on the type of experiment, and therefore the chosen protocol, we can measure from few to hundreds of oocytes daily. The importance of such a tool is illustrated by characterizing several variants of the Iml and ImII toxins. This new system provides a medium-throughput screening platform and expands by orders of magnitudes the number of samples and cells that can be measured in a single day while offering, in addition, the capacity to work with very limited sample size.

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Novel $\alpha 7$ nAChRs ligands: From virtual screening to functional assays

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Progresses made in the understanding of the tridimensional structure of the nicotinic acetylcholine receptors (nAChRs) allows to exploit new strategies for the finding of novel molecules acting at this class of ligand gated channels. Taking advantage of the crystal structure of the acetylcholine binding protein (AChBP), in silico ligand design from the chemical universe data base (GDB) [1] and virtual screening was performed. This allowed the identification of novel molecules that should display selectivity for the $\alpha 7$ nAChRs. Selected virtual hits were synthesized and their functional properties assessed at human nAChRs expressed in *Xenopus* oocytes. Experimental protocols were designed to probe the putative agonist or antagonist activities of these molecules. 72,945 virtual ligands were investigated using docking (Autodock and Glide) and shape similarity to known $\alpha 7$ ligands (ROCS). Thirty-eight structures among the 10% top-scoring virtual hits were selected for their structural novelty and ease of synthesis, prepared by standard organic synthesis methods, and their properties analyzed. Most of the molecules displayed antagonist properties with IC₅₀'s in the low micromolar range. Thus while this strategy properly identifies ligands that interact with the receptor, further refinement of our model is required for the identification of selective agonists. Altogether these data illustrate the power of combining virtual chemistry, small scale synthesis and electrophysiological approaches.

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Reference

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1.21

Homology models of the $\alpha 7$ acetylcholine receptor based upon bacterial receptors: Comparison of experimental and in silico derived data

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Unraveling the mechanistic link between agonist binding and ion permeation in ligand-gated channels remains a challenge for modern biophysics. The recent high-resolution crystallization of